# pDRIVE5SEAP-hHSP70 

A plasmid with a ubiquitous human heat shock protein 70 promoter<br>Catalog \# pDRIVE5s-hhsp70<br>For research use only<br>Version \# 11J04-MM

## PRODUCT INFORMATION

Content:

- 1 disk of lyophilized GT116 E. coli bacteria transformed by a pDRIVE5s plasmid.
- GT116 genotype is: $F$-, mcrA, $\Delta(m r r-h s d R M S-m c r B C)$, Ø80lacZ $\triangle M 15$, $\Delta l a c X 74, r s p L$ (StrA), recA1, endA1 $\Delta d c m \Delta s b c C-s b c D$.
- 4 pouches of E. coli Fast-Media Zeo (2 TB and 2 Agar).


## Shipping and storage:

- Products are shipped at room temperature.
- Transformed bacteria should be stored at $-20^{\circ} \mathrm{C}$. Bacteria are stable up to one year when properly stored.
- Store E. coli Fast-Media ${ }^{\circledR}$ Zeo at room temperature. Fast-Media ${ }^{\circledR}$ pouches are stable 18 months when stored properly.


## Quality control:

- Plasmid construct has been confirmed by restriction analysis and sequencing. - Bacteria have been lyophilized, and their viability upon resuspension has been verified.


## GENERAL PRODUCT USE

pDRIVE5s is an expression plasmid containing a native or composite promoter of interest. pDRIVE5s may be used to:

- Subclone a promoter of interest into another vector. Unique restriction sites are present at each end of the promoter allowing convenient excision. The 5' site is $S d a$ I. $S d a$ I is compatible with Nsi I and Pst I. The 3' restriction site is Nco I which includes the ATG start codon, and is compatible with $B s p H$ I and BspLU11 I.
- Compare the activity of different promoters in transient transfection experiments. Each pDRIVE5s promoter drives the expression of the SEAP reporter gene which allows for testing of the promoter's activity in transient transfection experiments. Furthermore, the SEAP gene is flanked by the restriction sites (Nco I and Nhe I) for easy replacement with a different gene of interest.


## PROMOTER CHARACTERISTICS

Heat Inducible Human Heat Shock Protein 70 (HSP70) promoter
Complete Promoter size: 465 bp
Specificity: Ubiquitous

The HSP70 gene encodes a major stress-inducible heat shock protein (HSP70) which plays an important role in protecting cells from deleterious stresses. Hsp70 is expressed constitutively in a wide variety of cells at normal growth temperatures. Its expression is induced by various physiological stresses, including exposure to heat shock and heavy metal, or under nonstress conditions, such as in response to serum stimulation. The HSP70 promoter contains at least two regulatory domains, a distal domain responsive to heat shock or heavy metals and a proximal domain responsive to stimulation by serum ${ }^{1}$. The activity of the HSP70 promoter can be induced by moderate hyperthermia $\left(39^{\circ} \mathrm{C} \text { to } 43^{\circ} \mathrm{C}\right)^{2}$, reaching expression levels similar to those of the CMV promoter.

1. Wu BJ., Kingston RE, Morimoto RI. 1986. Human HSP70 promoter contains at least two distinct regulatory domains. Proc Natl Acad Sci USA. 83(3):629-33.
2. Huang Q. et al. 2000. Heat-induced gene expression as a novel targeted cancer gene therapy strategy. Cancer Res. 60(13):3435-9.

## PLASMID FEATURES

- SEAP gene encodes an engineered secreted embryonic alkaline phosphatase. The levels of SEAP in the culture medium of transfected cells expressing the reporter gene can be assayed with chromogenic or luminescent methods
- SV40 pAn: The Simian Virus 40 late polyadenylation signal enables efficient cleavage and polyadenylation reactions resulting in high levels of steady-state mRNA.
- pMB1 Ori is a minimal E. coli origin of replication with the same activity as the longer Ori.
- EM2K is a bacterial promoter that enables the constitutive expression of the antibiotic resistance gene in E. coli.
- Sh ble gene confers Zeocin ${ }^{\mathrm{TM}}$ resistance therefore allowing the selection of transformed E. coli carrying a pDRIVE5s plasmid.
Note: Stable transfection of clones cannot be performed due to the absence of an eukaryotic promoter upstream of the Sh ble gene.


## METHODS

## Growth of pDRIVE5s-transformed bacteria:

Use sterile conditions to do the following:
1 - Resuspend the lyophilized $E$. coli by adding 1 ml of LB medium in the tube containing the disk. Let sit for 5 minutes. Mix gently by inverting the tube several times.
2- Streak bacteria taken from this suspension on a Zeocin ${ }^{\text {TM }}$ LB agar plate prepared with the E. coli Fast-Media ${ }^{\circledR}$ Zeo agar provided (see below).
3- Place the plate in an incubator at $37^{\circ} \mathrm{C}$ overnight.
4- Isolate a single colony and grow the bacteria in TB supplemented with zeocin using the Fast-Media ${ }^{\circledR}$ Zeo liquid provided (see below).
5- Extract the pDRIVE5s plasmid DNA using the method of your choice.

## Selection of bacteria with $E$. coli Fast-Media Zeo:

E. coli Fast-Media ${ }^{*}$ Zeo is a new, fast and convenient way to prepare liquid and solid media for bacterial culture by using only a microwave. E. coli Fast-Media ${ }^{\circledR}$ Zeo is a TB (liquid) or LB (solid) based medium with zeocin, and contains stabilizers.
E. coli Fast-Media ${ }^{\text {® }}$ Zeo can be ordered separately (catalog code fas-zn-1, fas-zn-s).

## Method:

1- Pour the contents of a pouch into a clean borosilicate glass bottle or flask.
2- Add 200 ml of distilled water to the flask
3- Heat in a microwave on MEDIUM power setting (about 400Watts), until bubbles start appearing (approximately 3 minutes). Do not heat a closed container. Do not autoclave Fast-Media ${ }^{\circledR}$.
4- Swirl gently to mix the preparation. Be careful, the bottle and media are hot, use heatproof pads or gloves and care when handling.
5- Reheat the media for 30 seconds and gently swirl again. Repeat as necessary to completely dissolve the powder into solution. But be careful to avoid overboiling and volume loss.
6- Let agar medium cool to $45^{\circ} \mathrm{C}$ before pouring plates. Let liquid media cool to $37^{\circ} \mathrm{C}$ before seeding bacteria.
Note: Do not reheat solidified Fast-Media ${ }^{\circledR}$ as the antibiotic will be permanently destroyed by the procedure.


EcoRI (23)
NotI (2) XbaI (19) SdaI (38)

1 GCGGCCGCGTCGACGATATCTAGAATTCGGATCCTGCAGGGCCGCCCACTCCCCCTTCCTCTCAGGGTCCCTGTCCCCTCCAGTGAATCCCAGAAGACTC
101 TGGAGAGTTCTGAGCAGGGGGCGGCACTCTGGCCTCTGATTGGTCCAAGGAAGGCTGGGGGGCAGGACGGGAGGCGAAAACCCTGGAATATTCCCGACCT

201 GGCAGCCTCATCGAGCTCGGTGATTGGCTCAGAAGGGAAAAGGCGGGTCTCCGTGACGACTTATAAAAGCCCAGGGGCAAGCGGTCCGGATAACGGCTAG

301 CCTGAGGAGCTGCTGCGACAGTCCACTACCTTTTTCGAGAGTGACTCCCGTTGTCCCAAGGCTTCCCAGAGCGAACCTGTGCGGCTGCAGGCACCGGCGC
401 GTCGAGTTTCCGGCGTCCGGAAGGACCGAGCTCTTCTCGCGGATCCAGTGTTCCGTTTCCAGCCCCCAATCTCAGAGCGGAGCCGACAGAGAGCAGGGAA

## NcoI (503)

501 CCCCATGGTTCTGGGGCCCTGCATGCTGCTGCTGCTGCTGCTGCTGGGCCTGAGGCTACAGCTCTCCCTGGGCATCATCCCAGTTGAGGAGGAGAACCCG $\rightarrow \quad M \quad V \quad L \quad G \quad P \quad C \quad M \quad L \quad L \quad L \quad L \quad L \quad L \quad L \quad G \quad L \quad R \quad L \quad Q \quad L \quad S \quad L \quad G \quad I \quad I \quad P \quad V \quad E \quad E \quad E \quad N \quad P$ 601 GACTTCTGGAACCGCGAGGCAGCCGAGGCCCTGGGTGCCGCCAAGAAGCTGCAGCCTGCACAGACAGCCGCCAAGAACCTCATCATCTTCCTGGGCGATG


701 GGATGGGGGTGTCTACGGTGACAGCTGCCAGGATCCTAAAAGGGCAGAAGAAGGACAAACTGGGGCCTGAGATACCCCTGGCTATGGACCGCTTCCCATA
 801 TGTGGCTCTGTCCAAGACATACAATGTAGACAAACATGTGCCAGACAGTGGAGCCACAGCCACGGCCTACCTGTGCGGGGTCAAGGGCAACTTCCAGACC
 901 ATTGGCTTGAGTGCAGCCGCCCGCTTTAACCAGTGCAACACGACACGCGGCAACGAGGTCATCTCCGTGATGAATCGGGCCAAGAAAGCAGGGAAGTCAG
 1001 TGGGAGTGGTAACCACCACACGAGTGCAGCACGCCTCGCCAGCCGGCACCTACGCCCACACGGTGAACCGCAACTGGTACTCGGACGCCGACGTGCCTGC
 1101 CTCGGCCCGCCAGGAGGGGTGCCAGGACATCGCTACGCAGCTCATCTCCAACATGGACATTGATGTGATCCTGGGTGGAGGCCGAAAGTACATGTTTCGC 199* S A $\mathrm{A} \quad \mathrm{R}$ 1201 ATGGGAACCCCAGACCCTGAGTACCCAGATGACTACAGCCAAGGTGGGACCAGGCTGGACGGGAAGAATCTGGTGCAGGAATGGCTGGCGAAGCGCCAGG
 1301 GTGCCCGGTATGTGTGGAACCGCACTGAGCTCATGCAGGCTTCCCTGGACCCGTCTGTGACCCATCTCATGGGTCTCTTTGAGCCTGGAGACATGAAATA
 SacII (1485)
1401 CGAGATCCACCGAGACTCCACACTGGACCCCTCCCTGATGGAGATGACAGAGGCTGCCCTGCGCCTGCTGAGCAGGAACCCCCGCGGCTTCTTCCTCTTC
 1501 GTGGAGGGTGGTCGCATCGACCACGGTCATCACGAAAGCAGGGCTTACCGGGCACTGACTGAGACGATCATGTTCGACGACGCCATTGAGAGGGCGGGCC 333* V 1601 AGCTCACCAGCGAGGAGGACACGCTGAGCCTCGTCACTGCCGACCACTCCCACGTCTTCTCCTTCGGAGGCTACCCCCTGCGAGGGAGCTCCATCTTCGG 366. Q L T S E E D D T L $\mathrm{S} \quad \mathrm{L} \quad \mathrm{V}$ 1701 GCTGGCCCCTGGCAAGGCCCGGGACAGGAAGGCCTACACGGTCCTCCTATACGGAAACGGTCCAGGCTATGTGCTCAAGGACGGCGCCCGGCCGGATGTT
 1801 ACCGAGAGCGAGAGCGGGAGCCCCGAGTATCGGCAGCAGTCAGCAGTGCCCCTGGACGAAGAGACCCACGCAGGCGAGGACGTGGCGGTGTTCGCGCGCG
 1901 GCCCGCAGGCGCACCTGGTTCACGGCGTGCAGGAGCAGACCTTCATAGCGCACGTCATGGCCTTCGCCGCCTGCCTGGAGCCCTACACCGCCTGCGACCT
 NheI (2069)
2001 GGCGCCCCCCGCCGGCACCACCGACGCCGCGCACCCGGGGCGGTCCCGGTCCAAGCGTCTGGATTGAAGCTAGCTGGCCAGACATGATAAGATACATTGA

2101 TGAGTTTGGACAAACCACAACTAGAATGCAGTGAAAAAAATGCTTTATTTGTGAAATTTGTGATGCTATTGCTTTATTTGTAACCATTATAAGCTGCAAT

## MfeI (2218)

2201 AAACAAGTTAACAACAACAATTGCATTCATTTTATGTTTCAGGTTCAGGGGGAGGTGTGGGAGGTTTTTTAAAGCAAGTAAAACCTCTACAAATGTGGTA
2301 TGGAATTAATTCTAAAATACAGCATAGCAAAACTTTAACCTCCAAATCAAGCCTCTACTTGAATCCTTTTCTGAGGGATGAATAAGGCATAGGCATCAGG $\rightarrow$
2401 GGCTGTTGCCAATGTGCATTAGCTGTTTGCAGCCTCACCTTCTTTCATGGAGTTTAAGATATAGTGTATTTTCCCAAGGTTTGAACTAGCTCTTCATTTC
2501 TTTATGTTTTAAATGCACTGACCTCCCACATTCCCTTTTTAGTAAAATATTCAGAAATAATTTAAATACATCATTGCAATGAAAATAAATGTTTTTTATT 2601 AGGCAGAATCCAGATGCTCAAGGCCCTTCATAATATCCCCCAGTTTAGTAGTTGGACTTAGGGAACAAAGGAACCTTTAATAGAAATTGGACAGCAAGAA 2701 AGCGAGCTTCTAGCTTATCCTCAGTCCTGCTCCTCTGCCACAAAGTGCACGCAGTTGCCGGCCGGGTCGCGCAGGGCGAACTCCCGCCCCCACGGCTGCT
 2801 CGCCGATCTCGGTCATGGCCGGCCCGGAGGCGTCCCGGAAGTTCGTGGACACGACCTCCGACCACTCGGCGTACAGCTCGTCCAGGCCGCGCACCCACAC
 SgrAI (2976)
2901 CCAGGCCAGGGTGTTGTCCGGCACCACCTGGTCCTGGACCGCGCTGATGAACAGGGTCACGTCGTCCCGGACCACACCGGCGAAGTCGTCCTCCACGAAG

3001 TCCCGGGAGAACCCGAGCCGGTCGGTCCAGAACTCGACCGCTCCGGCGACGTCGCGCGCGGTGAGCACCGGAACGGCACTGGTCAACTTGGCCATGATGG
 MfeI (3139)
3101 CTCCTCCTGTCAGGAGAGGAAAGAGAAGAAGGTTAGTACAATTGCTATAGTGAGTTGTATTATACTATGCAGATATACTATGCCAATGATTAATTGTCAA
3201 ACTAGGGCTGCAGGTTAATTAAGAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCG
3301 CCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTC
3401 GTGCGCTCTCCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCATAGCTCACGCTGTAGGTATC
3501 TCAGTTCGGTGTAGGTCGTTCGCTCCAAGCTGGGCTGTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTC
3601 CAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGG
3701 TGGCCTAACTACGGCTACACTAGAAGAACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCA

3801 AACAAACCACCGCTGGTAGCGGTGGTTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTTCTACGGG
3901 GTCTGACGCTCAGTGGAACGAAAACTCACGTTAAGGGATTTTGGTCATGGCTAGTTAATTAACATTTAAATCA

# Fast-Media ${ }^{\circledR}$ 

# Microwaveable media for selection and propagation of E. coli transformants 

Catalog \# fas-xx-1, fas-xx-s, fas-xx-xgal
For research use only
Version \# 10G07-MM

## PRODUCT INFORMATION

## Contents:

E. coli Fast-Media ${ }^{\circledR}$ are prepared as individual sealed pouches containing the necessary amount of powder for preparation of 200 ml of selective liquid or agar medium.
30 pouches are supplied for each order of TB or Agar and 20 pouches are supplied for each order of XGal Agar.
Storage and stability:
Fast-Media ${ }^{\circledR}$ are shipped at room temperature, and must be stored in a dry and cool place. They are stable for 2 years at room temperature.
When properly prepared, Fast-Media ${ }^{\star}$ plates or broths are stable for 4 weeks at $4^{\circ} \mathrm{C}$, and remain sterile and selective.

## Quality control:

The high quality and performance of each formulation has been tested with some widely used and proprietary E. coli K12 derived strains*. These include DH5 $\alpha$, Top10, MC1061, XL1 blue, JM 109, TB1, GT100, GT110, GT115, GT116.
The adequate plasmids carrying the appropriate E. coli resistance genes are used as positive control.
*E. coli recipient strains carrying the Tn 5 transposon are resistant to Kanamycin and Zeocin"'.

## GENERAL PRODUCT USE

E. coli Fast-Media ${ }^{\text {® }}$ are microwaveable ready-to-use solid or liquid media, supplied with a selective antibiotic, and chromogenic substrates (for five references), therefore designed for the growth or selection of E. coli transformant colonies, as well as detection of blue/white colonies.

- Fast-Media ${ }^{\text {® }}$ Agar formulation is LB based agar medium supplemented with selective antibiotic, it is used for selection of resistant $E$. coli colonies after transformation by vectors carrying a selection resistance gene.
- Fast-Media ${ }^{\otimes} \mathbf{X}$-Gal formulation is a LB based agar medium supplemented with selective antibiotic, X-Gal and IPTG. It is used for detection of blue/white resistant colonies after transformation by a vector carrying $\operatorname{Lac} Z$ gene.
- Fast-Media ${ }^{\circledR}$ TB formulation is a Terrific Broth based liquid medium supplemented with selective antibiotic. It's used for high cell density culture of transformed bacteria, and extraction of high quantity and quality of required plasmid.


## FAST-MEDIA ${ }^{\circledR}$ FEATURES

E. coli Fast-Media ${ }^{\text {® }}$ offer researchers a quick and convenient way to prepare 200 ml of liquid culture medium, or 8-10 agar plates in about five minutes USING A MICROWAVE INSTEAD OF AN AUTOCLAVE.
E. coli Fast-Media ${ }^{\text {® }}$ are available with a large variety of prokaryotic selective agents including Ampicillin, Blasticidin S, Hygromycin B, Kanamycin, Puromycin and Zeocin ${ }^{\text {" }}$ (see table below). Fast-Media ${ }^{\text {® }}$ is also available with no selective agent (Base) that can be prepared with or without antibiotics.

|  | Agar | X-Gal | TB |
| :---: | :---: | :---: | :---: |
| Base | $\checkmark$ |  | $\checkmark$ |
| Ampicillin | $\checkmark$ | $\checkmark$ | $\checkmark$ |
| Blasticidin | $\sqrt{ }$ | $\sqrt{ }$ | $\checkmark$ |
| Hygromycin | $\sqrt{ }$ | $\sqrt{ }$ | $\sqrt{ }$ |
| Kanamycin | $\checkmark$ | $\checkmark$ | $\checkmark$ |
| Puromycin | $\sqrt{ }$ |  | $\sqrt{ }$ |
| Zeocin'" | $\checkmark$ | $\checkmark$ | $\checkmark$ |

## SPECIAL HANDLING

Caution should be exercised during handling of Fast-Media ${ }^{\circledR}$ due to potential allergenic properties of antibiotics. Wear protective gloves, do not breath the dust.

## METHOD

For customer convenience, procedure is directly printed on each pouch.
1- Pour the pouch contents into a clean borosilicate glass bottle or flask.
2- Add 200 ml of distilled or deionized water.
3- Mix thoroughly by swirling the glass bottle or flask.
4- Heat in a microwave oven on MEDIUM power setting (about 450W) until bubbles start to appear (about 3 minutes).

## Do not heat in a closed container.

5- Swirl gently to mix the preparation and re-heat for 30 seconds. Swirl gently again.
6- Repeat step 4 if necessary until the medium is completely dissolved. Do not overboil.
7- Allow the medium to cool to $50-55^{\circ} \mathrm{C}$, use directly for liquid medium, or pour plates for solid medium.
Caution: Any solution heated in a microwave oven may become superheated and suddenly boil when moved or touched. Handle with extreme care. Wear heat-proof gloves.
Note: Do not repeat this above procedure once the medium is prepared because the antibiotic will be adversely affected.

## For preparation of supplemented Fast-Media ${ }^{\circledR}$ Base.

- Follow the instructions above and when media has cooled to $50-55^{\circ} \mathrm{C}$ add the antibiotic at the appropriate concentration for selection of E. coli.

